



Novel vitamin D receptor ligands having a carboxyl group as an anchor to arginine 274 in the ligand-binding domain[☆]

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ABSTRACT

Vitamin D₃ is metabolized into the hormonally active form, 1 α ,25-dihydroxyvitamin D₃ (**1**), via 25-hydroxyvitamin D₃ (**2**) which is the most abundant circulating metabolite. Introduction of the 1 α -hydroxyl group into 25-hydroxyvitamin D₃ (**2**) to produce 1 α ,25-dihydroxyvitamin D₃ (**1**) increases the VDR binding affinity by approximately 1000-fold. The X-ray crystal structure of human VDR in complex with 1 α ,25-dihydroxyvitamin D₃ (**1**) shows that, together with Ser-237, the 1 α -hydroxyl group of 1 α ,25-dihydroxyvitamin D₃ (**1**) makes hydrogen bonds with Arg-274, single mutation of which results in impaired ligand recognition. In 2002, lithocholic acid, which possesses a carboxyl group at position C24, was demonstrated to be a weak VDR ligand. We speculated that the carboxylic acid of lithocholic acid could be recognized by Arg-274 in the ligand-binding domain of VDR. In view of the significance of Arg-274 to direct the 1 α -hydroxyl group, as well as the results with lithocholic acid and its derivatives, we designed the C2 modified analogues of 25-hydroxyvitamin D₃ (**2**) having a carboxyl group, instead of the 1-hydroxyl group, for better electrostatic interaction to the guanidinium side-chain of arginine.

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1. Introduction

Vitamin D₃ is metabolized into the hormonally active form, 1 α ,25-dihydroxyvitamin D₃ (**1**), via 25-hydroxyvitamin D₃ (**2**) which is the most abundant circulating metabolite (Fig. 1) [1–3]. Introduction of the 1 α -hydroxyl group into 25-hydroxyvitamin D₃ (**2**) to produce 1 α ,25-dihydroxyvitamin D₃ (**1**) increases the vitamin D receptor (VDR) binding affinity by approximately 1000-fold. Several enzymes, including CYP27A1 and CYP27B1, strictly regulate formation and degradation of the most active metabolite **1**, depending upon serum calcium concentration [4].

Most actions of **1** as well as its fellow metabolites are mediated via a ligand-inducible transcription factor, VDR, belonging to the nuclear receptor super-family. Binding mode of ligands to VDR, as well as the stability of the complex, is now a focus of interest for chemists, because formation of the ligand–receptor complex triggers the whole sequential reactions, in vitamin D-responsive gene activation and/or suppression, resulting in specific biological responses.

The X-ray crystal structures of human VDR in complex with **1** and its analogues show that each of the three hydroxyl groups at

positions C1, C3 and C25 makes hydrogen bonds with two amino acid residues in the ligand-binding domain (LBD) of VDR (Fig. 2) [5,6]. In particular, together with Ser-237, the 1 α -hydroxyl group in **1** makes hydrogen bonds with Arg-274, single mutation of which results in impaired ligand recognition [7]. In return, the importance of the 1 α -hydroxyl group over the other hydroxyl groups at position C3 and C25 in the *seco*-steroid structure has been demonstrated by the data of A-ring diastereomers of **1** [4,8,9]. In comparison with the case of the 3-hydroxyl group, epimerization or loss of the hydroxyl group at position C1 severely affects the VDR affinity.

In 2002, lithocholic acid (**3**: LCA) and 3-ketolithocholic acid (**4**), which possess a carboxyl group at position C24, were demonstrated to be weak VDR ligands [10]. Although acylation at the 3-hydroxyl group in **3** increased the potency, the transformation of the 24-carboxylic acid in LCA **3** into its methyl, ethyl or benzyl ester decreased the potency of the VDR activation, which suggested the importance of the free carboxylic acid to be recognized by VDR [11]. We envisioned that the carboxylic acid of **3**, instead of the hydroxyl group, could make interaction with the critical Arg-274 in the LBD of VDR. In view of the significance of Arg-274 to direct the 1 α -hydroxyl group in vitamin D compounds, as well as the results with lithocholic acid derivatives, we designed the C2 modified analogues of 25-hydroxyvitamin D₃ (**2**) having a carboxyl group, instead of the 1-hydroxyl group, for better electrostatic interaction to the guanidinium side-chain of arginine (Fig. 3). Novel VDR ligands, 2 α -carboxy-25-hydroxyvitamin D₃ (**5**) as well as its methyl ester (**6**), as a reference compound of **5**, have been synthesized (Fig. 4).

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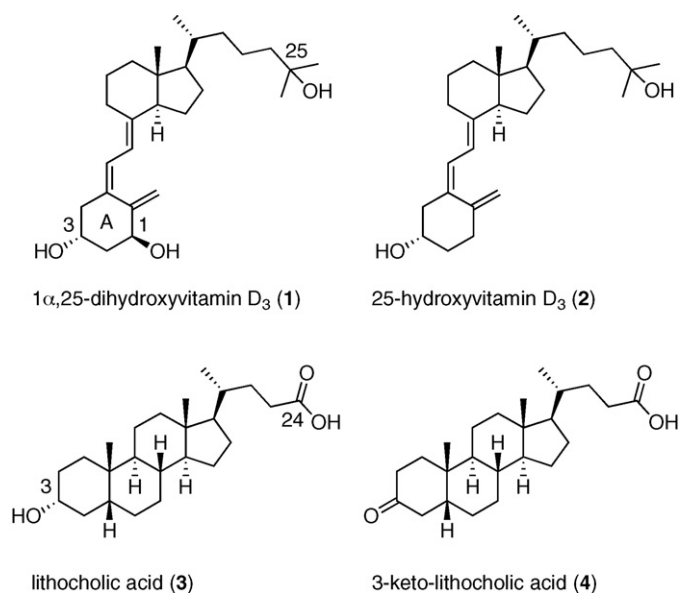


Fig. 1. Chemical structures of 1 α ,25-dihydroxyvitamin D₃ (1), 25-hydroxyvitamin D₃ (2), lithocholic acid (3) and 3-keto-lithocholic acid (4).

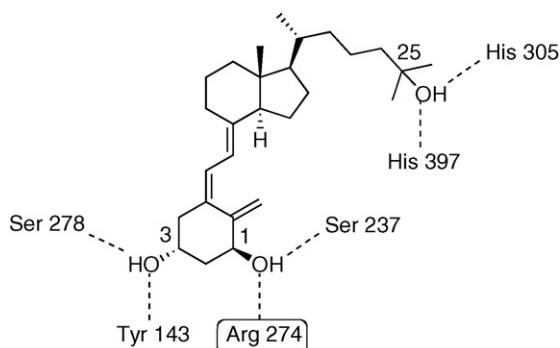


Fig. 2. Each of the three hydroxyl groups at positions C1, C3 and C25 in **1** was recognized with the two amino acid residues in the ligand-binding domain (LBD) of human VDR. The crucial 1 α -hydroxyl group in **1** made hydrogen bonds with Arg-274 and Ser-237.

Novel VDR ligands with a carboxyl group

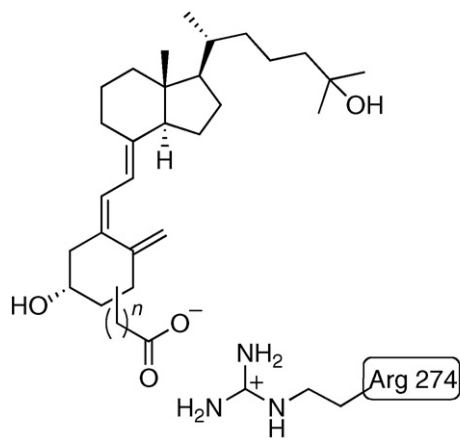


Fig. 3. Design of novel VDR ligands with a carboxyl group. The carboxyl group instead of the 1 α -hydroxyl group could interact the guanidinium side-chain of Arg-274 in the LBD more favourably.

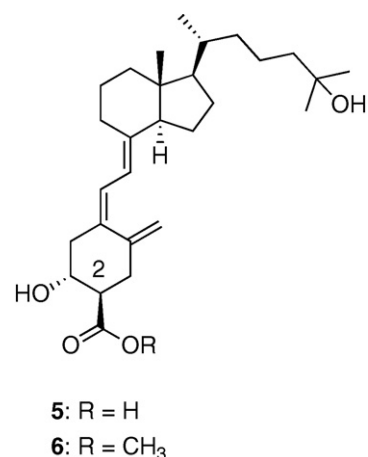
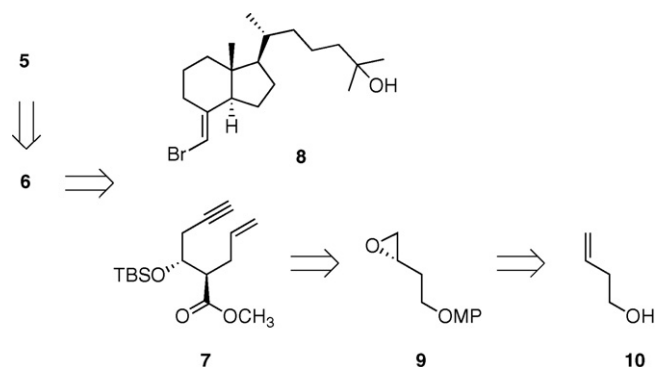


Fig. 4. Chemical structure of 2 α -carboxy-25-hydroxyvitamin D₃ (5) and its methyl ester (6).



Scheme 1. Retrosynthesis of 2 α -carboxy-25-hydroxyvitamin D₃ (5). MP: 4-methoxyphenyl.

2. Results and discussion

Synthesis of 2 α -carboxy-25-hydroxyvitamin D₃ (5) and its methyl ester (6) has been carried out by a convergent method using a palladium catalyst [12]. Scheme 1 depicts the retrosynthetic analysis of the novel VDR ligands (5, 6) (Scheme 1). Separate synthesis of the requisite A-ring enyne precursor 7 and the CD-ring portion 8, followed by coupling of the two units, produced the targeted compounds. Nine-step conversion of the known epoxide 9 prepared from 3-buten-1-ol 10 yielded the requisite A-ring enyne precursors of 6, which was then converted to 5 by hydrolysis. Detailed synthetic procedures will be published elsewhere soon.

Preliminary testing of the novel VDR ligands (5, 6) showed weak affinity to bovine thymus VDR in comparison with 1 α ,25-dihydroxyvitamin D₃ (1), however their potencies were comparable to 25-hydroxyvitamin D₃ (2). Under the conditions that we employed, lithocholic acid 3 itself exhibited low VDR affinity; our novel compounds showed more than 100-fold better binding to VDR in comparison with 3.

In summary, we have designed novel VDR ligands introduced a carboxyl group into the A-ring of the *seco*-steroid to interact the critical amino acid residue, Arg-274 in the LBD of VDR. Synthesis of the compounds was carried out by a convergent method using palladium catalyst. Novel VDR ligands showed comparable VDR binding affinities to VDR in comparison with 25-hydroxyvitamin D₃.

References

- [1] D. Feldman, J.W. Pike, F.H. Glorieux (Eds.), *Vitamin D*, second ed., Elsevier Academic Press, Burlington, 2005.
- [2] J. Reichrath, M. Friedrich, W. Tilgen (Eds.), *Vitamin D Analogs in Cancer Prevention and Therapy*, Springer-Verlag, Berlin, 2003.
- [3] M.F. Holick (Ed.), *Vitamin D: Physiology, Molecular Biology, and Clinical Applications*, Humana Press, Totowa, 1999.
- [4] R. Bouillon, W.H. Okamura, A.W. Norman, Structure–function relationships in the vitamin D endocrine system, *Endocr. Rev.* 16 (1995) 200–257.
- [5] N. Rochel, J.M. Wurtz, A. Mitschler, B. Klaholz, D. Moras, The crystal structure of the nuclear receptor for vitamin D bound to its natural ligand, *Mol. Cell* 5 (2000) 173–179.
- [6] S. Hourai, T. Fujishima, A. Kittaka, Y. Suhara, H. Takamaya, N. Rochel, D. Moras, Probing a water channel near the A-ring of the receptor bound $1\alpha,25$ -dihydroxyvitamin D_3 with selected 2α -substituted analogues, *J. Med. Chem.* 49 (2006) 5199–5205.
- [7] P.J. Malloy, J.W. Pike, D. Feldman, The vitamin D receptor and the syndrome of hereditary $1,25$ -dihydroxyvitamin D-resistant rickets, *Endocr. Rev.* 20 (1999) 156–188.
- [8] T. Fujishima, K. Konno, K. Nakagawa, M. Kurobe, T. Okano, H. Takayama, Efficient synthesis and biological evaluation of all A-ring diastereomers of $1\alpha,25$ -dihydroxyvitamin D_3 and its 20-epimer, *Bioorg. Med. Chem.* 8 (2000) 123–134.
- [9] K.R. Muralidharan, A.R. De Lera, S.D. Isaef, A.W. Norman, W.H. Okamura, Studies on the A-ring diastereomers of $1\alpha,25$ -dihydroxyvitamin D, *J. Org. Chem.* 58 (1993) 1895–1899.
- [10] M. Makishima, T.T. Liu, W. Xie, G.K. Whitfield, H. Domoto, R.M. Evans, M.R. Haussler, D.J. Mangelsdorf, Vitamin D receptor as an intestinal bile acid sensor, *Science* 296 (2002) 1313–1315.
- [11] R. Adachi, Y. Honma, H. Masuno, K. Kawana, I. Shimoura, S. Yamada, M. Makishima, Selective activation of vitamin D receptor by lithocholic acid acetate, a bile acid derivative, *J. Lipid Res.* 46 (2005) 46–57.
- [12] B.M. Trost, J. Dumas, M. Villa, New strategies for the synthesis of vitamin D metabolites via Pd-catalyzed reactions, *J. Am. Chem. Soc.* 114 (1992) 9836–9845.